

## REVIEW

# The roles of microRNAs in the progression of castration-resistant prostate cancer

Satoko Kojima<sup>1</sup>, Yusuke Goto<sup>2</sup> and Yukio Naya<sup>1</sup>

Prostate cancer (PCa) is one of the leading causes of cancer-related death in men. PCa is androgen-dependent, and androgen-deprivation therapy is effective for first-line hormonal treatment, but the androgen-independent phenotype of PCa eventually develops, which is difficult to treat and has no effective cure. Recently, microRNAs have been discovered to have important roles in the initiation and progression of PCa, suggesting their use in diagnosis, predicting prognosis and development of treatment for castration-resistant PCa (CRPC). Understanding the networks of microRNAs and their target genes is necessary to ascertain their roles and importance in the development and progression of PCa. This review summarizes the current knowledge about microRNAs regulating PCa progression and elucidates the mechanism of progression to CRPC.

*Journal of Human Genetics* (2017) 62, 25–31; doi:10.1038/jhg.2016.69; published online 9 June 2016

## INTRODUCTION

Prostate cancer (PCa) is the most frequent malignant tumor and the second leading cause of cancer death in men in Western countries.<sup>1</sup> Recently, multimodality treatments have become available, but the prognosis is still poor once a patient progresses to castration-resistant PCa (CRPC). PCa is initially androgen-sensitive, and androgen-deprivation therapy is widely used for metastatic PCa patients. The disease is known to progress to CRPC about 12 months after androgen-deprivation therapy.<sup>2</sup> As new hormonal therapies, abiraterone (CYP-17A inhibitor)<sup>3</sup> and enzalutamide (second-generation anti-androgen)<sup>4</sup> have been developed to treat CRPC, and cabazitaxel (second-generation chemotherapy)<sup>5</sup> has been developed for patients who relapse to docetaxel. However, the effects of these new therapies are limited, and patients progress to the lethal stage, with overall survival increasing only 3–4 months after becoming CRPC compared with controls. Prostate-specific antigen (PSA) is a highly specific marker used to detect PCa, but multiple factors including genetic variations, growth factors and androgen receptor (AR) status affect disease progression and prognosis.<sup>6</sup>

MicroRNAs (miRNAs) are a class of small non-coding RNAs that have important roles in cell development, differentiation, signal transduction, cancer formation and progression by regulating the expressions of protein-coding genes by repressing translation or cleaving of RNA transcripts in a sequence-specific manner.<sup>7</sup> In addition, miRNAs are known to have important roles in the regulation of malignant transformation and development of PCa.<sup>8</sup>

Recent clinical priorities include the identification of biomarkers that discriminate between low- and high-risk diseases to select appropriate treatment for each patient. From the profiling of expressions of miRNAs in PCa, many miRNAs are consistently upregulated

or downregulated, suggesting that certain biomarkers predict prognosis in PCa. It is important to determine the target genes of these differentially expressed miRNAs to elucidate their functions. The roles of miRNAs in PCa can be divided by their associations with cell proliferation, apoptosis, invasion and metastasis, epithelial–mesenchymal transition (EMT), cancer stemness, and AR status. First, we introduce the expression profiles of miRNAs in PCa, and we discuss miRNAs that have important effects on each categorized function in PCa. We also addressed possible therapeutic roles of miRNAs and significance of miRNAs as predictive biomarkers in CRPC.

## EXPRESSION PROFILES OF MIRNAS IN PCA

To date, 2578 human mature miRNAs have been registered in the public database (miRBase, <http://microrna.sanger.ac.uk/>, release 21 June 2014). A growing body of evidence suggests that expression profiles of miRNAs in PCa are increasing in importance because of their usefulness for diagnosis, staging, progression, and predicting prognosis and response to treatment.<sup>9</sup> Differential expressions of miRNAs in PCa were analyzed using current genome-based technologies, which have been able to distinguish benign prostate tissue from PCa. Recently, miRNA signatures comparing expressions of miRNAs in benign prostate tissue and CRPC clinical specimens obtained from autopsy have been published.<sup>10</sup> Upregulation of *miR-96*, *-182*, *182\**, *-183*, *-375*, *32*, *-26a*, *-181a*, *-93*, *-196a*, *-25*, *-92* and *let-7i* and downregulation of *miR-16*, *-31*, *-125b*, *-145*, *-149*, *-181b*, *-184*, *-205*, *-221* and *-222* were confirmed in PCa tissue.<sup>11,12</sup> Many other biomarkers have been reported using hormone-sensitive PCa<sup>13</sup> or CRPC<sup>10,14,15</sup> and serum or urine of PCa patients.<sup>16,17</sup> It appears that miRNAs may function as oncogenes or tumor suppressors. Oncogenic miRNAs are upregulated, and tumor-suppressive miRNAs are

<sup>1</sup>Department of Urology, Teikyo University Chiba Medical Center, Ichihara, Japan and <sup>2</sup>Department of Functional Genomics, Chiba University Graduate School of Medicine, Chiba, Japan

Correspondence: Dr S Kojima, Department of Urology, Teikyo University Chiba Medical Center, 3426-3 Anesaki, Ichihara 299-0111, Japan.

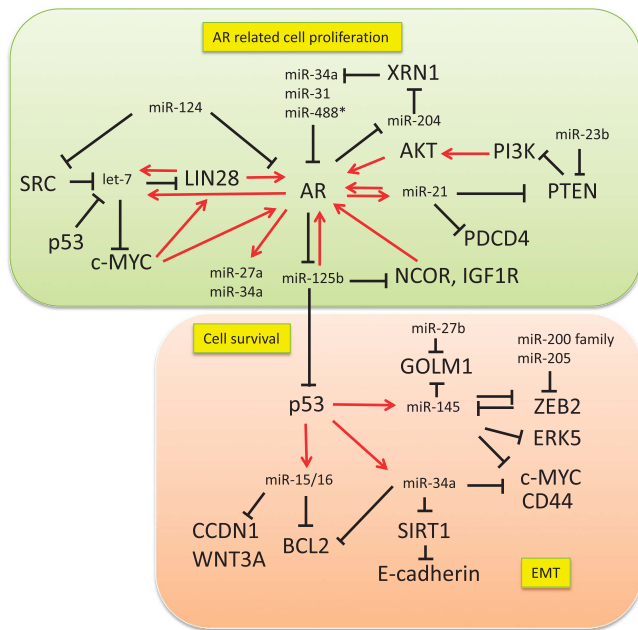
E-mail: s-kojima@med.teikyo-u.ac.jp

Received 20 March 2016; revised 28 April 2016; accepted 10 May 2016; published online 9 June 2016

**Table 1 Expression, gene location, target genes and functions of each miRNA in prostate cancer**

miRNAs	Expression	Chromosomal location	Targets in PCa	Functions and key findings
let-7a-1,2,3	Down	9q22.32, 11q24.1, 22q13.31	E2F2, <sup>100</sup> CCND2, <sup>100</sup> IGF1R, <sup>101</sup> EZH2 <sup>59</sup>	Inhibits cell proliferation and cell growth
let-7-b	Down	22q13.31	EZH2 <sup>59</sup>	Inhibits cell proliferation and cell growth
let-7c	Down	21q21.1	c-MYC, <sup>62</sup> Lin28, <sup>62</sup> EZH2, <sup>59</sup> IGF1R <sup>23</sup>	AR antagonize
miR-125b-1	Up	11q24.1	P53, <sup>21</sup> BAK-1, <sup>21,102</sup> Puma, <sup>21</sup> NCoR, <sup>68</sup> E1F4EBP1, <sup>46</sup> IGF1R <sup>23</sup>	Increases cell proliferation and inhibits apoptosis, activates AR signaling
miR-125b-2	Up	21q21.1		
miR-133a-1	Down	18q11.2	EGFR, <sup>103</sup> PNP <sup>104</sup>	Inhibits cell proliferation, invasion and migration
miR-133a-2	Down	20q13.33		
miR-143/145	Down	5q32	KRAS, <sup>105</sup> ERK5, <sup>106</sup> FSCN1, <sup>48</sup> SWAP70, <sup>49</sup> MMP13, KLKs, <sup>107</sup> GOLM1, <sup>52</sup> CD133, <sup>50</sup> CD44, <sup>50</sup> MYC, <sup>50</sup> KLF4, <sup>50</sup> OCT4, <sup>50</sup> SOX2, <sup>108</sup> LMK1, <sup>109</sup> DAB2, <sup>110</sup> ZEB2, <sup>51</sup> ERG <sup>111</sup>	Inhibits cell proliferation, invasion and migration, reversing EMT
miR-15/16-1	Down	13q14.2	BCL2, <sup>25</sup> CCND1, <sup>25</sup> WNT3A, <sup>25</sup> CDK1, <sup>26</sup> CDK2, <sup>26</sup> FGF-2, <sup>112</sup> FGFR1 <sup>112</sup>	Inhibits cell proliferation, invasion and migration, angiogenesis
miR-16-2	Down	3q25.33		
miR-200a, 200b	Down	1q36.33	ZEB1, <sup>113</sup> ZEB2, <sup>113</sup> SNAIL2, <sup>113</sup> SLUG <sup>114</sup>	Reversing EMT, <sup>115</sup> docetaxel-resistant <sup>90</sup>
miR-200c	Down	12p13.31		
miR-205	Down	1q32.2	c-SRC, <sup>116</sup> ErbB3, <sup>117</sup> E2F1, <sup>117</sup> ZEB2, <sup>117</sup> BCL-w, <sup>118</sup> E2F6, <sup>118</sup> MAPK and AR signaling, <sup>119</sup> TP53INP1 <sup>120</sup>	Inhibits cell proliferation, invasion, migration and reversing EMT, AR signaling, docetaxel-resistant <sup>118</sup>
miR-21	Up	17q23.2	MARCKS, <sup>36</sup> RECK, <sup>121</sup> PTEN, <sup>32</sup> PDCD4 <sup>33</sup>	Increases cell migration and invasion; EMT, docetaxel-resistant <sup>63</sup>
miR-221/222	Down	Xp11.3	Ecm29, <sup>10</sup> SIRT1, <sup>122</sup> IRF2, <sup>123</sup> SOCS3, <sup>123</sup> IRF2, <sup>123</sup> Bmi-1 <sup>124</sup>	Inhibits cell proliferation, invasion and migration
miR-23b	Down	9q22.32	PTEN, <sup>29</sup> Scr kinase, <sup>125</sup> MAPK and JAK/STAT pathway <sup>126</sup>	Proliferation, invasion, migration
miR-34a	Down	1p36.22	AR, <sup>127</sup> NOTCH1 signaling, SIRT1, <sup>128</sup> CD44, <sup>57</sup> LEF1, <sup>129</sup>	Stimulates cancer stemness, tumorigenesis, proliferation, migration, AR regulation, associated with GS, LN metastasis
miR-34b, 34c	Down	11q23.1	c-MYC, <sup>130</sup> BCL2, HuR <sup>131</sup>	
miR-488*	Down	1q25.2	AR <sup>66</sup>	Inhibits AR-mediated cell growth

Abbreviations: AR, androgen receptor; EMT, epithelial-mesenchymal transition; GS, gleason score; LN, lymph nodes.



**Figure 1** Physiological roles of miRNAs in prostate cancer. miRNA targets and functions are categorized into relation with androgen receptor, cell survival and EMT.

downregulated in cancers. The role of miRNAs in PCa is understood by elucidating the relationships of miRNAs and their target genes. The miRNAs that have been shown to have important functions in PCa and whose target genes have been determined are listed in Table 1, and the association of each miRNA and the target genes are shown in Figure 1.

### MIRNAS ASSOCIATED WITH CELL PROLIFERATION AND APOPTOSIS IN PCA

*p53* is a tumor suppressor gene, and loss of *p53* function has a critical role in the development of PCa.<sup>18,19</sup> *p53* mutation is a late event in the progression of PCa and is associated with advanced stage, loss of differentiation and the transition from androgen-dependent to androgen-independent growth.<sup>20</sup> Shi et al.<sup>21</sup> found that *miR-125b*, which is aberrantly expressed in PCa cells and tissues, promoted the growth of PCa xenograft through downregulation of three key pro-apoptotic genes, *p53*, *p53* upregulated modulator of apoptosis (*PUMA*) and *Bak1*. Thus, increased expression of onco-*miR-125b* decreased *p53* expression, resulting in survival of PCa cells. Insulin-like growth factor 1 receptor (*IGF1R*) is important for the tumorigenesis and progression of PCa because of its demonstrated roles in angiogenesis, transformation and mitogenesis.<sup>22</sup> Recently, it has been reported that *miR-99a/let-7c/125b* cluster, which is transcriptionally repressed by androgen-activated AR, has been shown to directly target *IGF1R* in androgen-dependent PCa cells.<sup>23</sup> This might be one of the mechanisms by which AR induces cell proliferation in PCa.

Two miRNAs, *miR-15a* and *miR-16-1*, are located at ch13q14, a locus in which complete or partial genomic loss is reported in advanced PCa and is associated with tumor initiation, progression and metastasis.<sup>24</sup> Downregulation of these miRNAs is significant in advanced PCa.<sup>25</sup> The *miR-15a-miR-16-1* cluster has been reported to regulate various oncogenes, such as B-cell lymphoma 2 (*BCL2*), cyclin D1 (*CCND1*) and wingless-related MMTV integration site 3A (*WNT3A*), through posttranscriptional repression in PCa.<sup>25</sup>

Deregulated expression of Bcl2 and cyclins is commonly reported in PCa, which is thought to facilitate the survival of cells under androgen-depletion therapy. Overexpression of *miR-16* in PCa cell lines significantly inhibits the growth of prostate tumors through the downregulation of CDK1 and CDK2 in bone, suggesting that *miR-16* could represent a novel type of personalized therapy for treating metastatic PCa.<sup>26</sup>

Monoallelic loss or mutation of phosphatase and tensin homolog (*PTEN*) is detected in the early stages of many sporadic tumors, including PCa.<sup>27</sup> Downregulation of *PTEN*, allowing activation of phosphatidylinositol 3-kinase (PI3K)-AKT pathway, results in decreased cell apoptosis and provision of cell survival signals.<sup>28</sup> Recently, studies have increased to investigate the roles of miRNAs in *PTEN* regulation. *miR-19b*, *miR-23b*, *miR-26a* and *miR-92a* have been reported to promote prostate cell proliferation by targeting *PTEN* and inhibiting the PI3K/AKT pathway, and cyclin D1 *in vitro*.<sup>29</sup> Other studies have also confirmed that *miR-22* and the *miR-106b~25* cluster are either directly or indirectly involved in the *PTEN* regulation in PCa.<sup>30</sup>

It has been shown that *miR-21* is one of the most common deregulated oncomiRs, which has an important role in cancer pathogenesis, and high expression of *miR-21* is found in almost all types of solid cancer tissues including PCa.<sup>31</sup> Furthermore, *miR-21* directly targets tumor suppressor genes such as *PTEN*<sup>32</sup> and programmed cell death 4 (*PDCD4*)<sup>33</sup> in PCa, and *miR-21* is induced by AR with binding to the promoter site of *miR-21*, resulting in the overexpression of *miR-21*, leading to the castration resistance phenotype of PCa.<sup>34,35</sup> Clinically, *miR-21* is a useful prognostic marker, which is associated with PCa recurrence after radical prostatectomy.<sup>36,37</sup>

A cell line-based genomic approach showed that the *miR-221/222* cluster was upregulated in LNCaP cells in the castration-resistant condition and was reported as oncogenic, promoting metastasis of PCa.<sup>38</sup> One of the mechanisms of *miR-221/222* on tumor cell proliferation in PCa cell lines is directly targeting *p27kip1*, a tumor suppressor gene.<sup>39</sup> The *miR-221/222* cluster is also reported to promote cell proliferation and repress apoptosis thorough suppressing caspase-10.<sup>40</sup> However, genome-wide miRNA expression signatures using clinical PCa and CRPC specimens showed significant downregulation of the *miR-221/222* cluster compared with normal prostate tissues, suggesting these miRNAs function as tumor suppressors in PCa patients, especially in CRPC.<sup>10,12,13,41–45</sup> These comprehensive approaches to miRNA expression in clinical specimens would help identify novel functional mechanisms of miRNAs.

### MIRNAS ASSOCIATED WITH CELL MIGRATION, INVASION, EMT AND STEMNESS IN PCA

It was previously reported that *miR-145* was downregulated in many solid tumors, including PCa and functioned as a tumor suppressor,<sup>15,46</sup> and that *miR-145* was upregulated by wild-type *p53*.<sup>47</sup> *miR-145* directly targets fascin homolog 1 (*FSCN1*)<sup>48</sup> and switching B-cell complex 70kDa subunit (*SWAP70*).<sup>49</sup> Knockdown of *FSCN1* and *SWAP70* suppresses cell migration and invasion in PCa cells. Restoration of *miR-145* inhibits cell proliferation in PCa. The *miR-143/145* cluster is downregulated in PCa cells, resulting in enhanced cell migration and invasion through reduced expression of E-cadherin, thus promoting the EMT phenotype.<sup>50,51</sup> *miR-145* targets zinc-finger E-box binding homeobox 2 (*ZEB2*), as an EMT activator, and *ZEB2* directly represses the transcription of *miR-145*. This double-negative feedback loop has important roles in suppressing tumor cell

invasion, migration and EMT.<sup>51</sup> Golgi membrane protein 1 (*GOLM1*) has also been identified as a target of *miR-143/145*, and it is considered to have roles in tumor cell proliferation, migration and invasion.<sup>52</sup> Moreover, another tumor-suppressive miRNA, *miR-27b*, regulates *GOLM1*, which indicates that several tumor-suppressive miRNAs regulate *GOLM1* in concert.<sup>53</sup>

Cancer stem cells (CSCs) are a subset of cancer cells that have important roles in tumor progression and metastasis in several cancers, including PCa.<sup>54</sup> *CD44*, an adhesion molecule, is a marker to identify CSCs, and the expression of *CD44* was found to be increased in a cell population with increased potential for tumor initiation and metastasis.<sup>55</sup> Studies have clearly shown that miRs are involved in promoting or inhibiting the stemness of CSCs;<sup>56</sup> *miR-143* and *miR-145* suppress tumor sphere formation and expression of CSC markers and stemness factors, including *CD133*, *CD44*, *Oct4*, *c-Myc* and *Klf4* in PC3 cells,<sup>50</sup> indicating that *miR-143* and *miR-145* may have a significant role in bone metastasis progression of PCa by regulating CSC characteristics.<sup>50</sup>

*miR-34a* has been shown to be downregulated in CD44+ PCa cells, and these cells have increased inhibition of clonogenic growth, metastatic behavior and tumor regenerator.<sup>57,58</sup> The tumor suppressor *p53* induces transcription of *miR-34a*, which is known to have strong anti-tumor effects. The *let-7* family also appears to have a key role in the recurrence and progression of PCa by regulating CSCs. The *let-7* family was found to be lost in PCa tissue specimens with Gleason score 7 or higher, with consistently increased expression of Enhancer of Zeste homolog 2 (*EZH2*).<sup>59</sup> *EZH2* is a putative target of the *let-7* family and was demonstrated to control stem cell function in PCa cells.<sup>59</sup>

As for neuroendocrine differentiation (NED), the *AR-miR-204-XRN1* (5'-3' exoribonuclease 1) axis has been reported to contribute to NED. *miR-204* functions as a tumor suppressor in AR-positive LNCaP and 22Rv1 cells, but as an oncogene in PC3 and CL1 cells, and these dual functions of miRNAs provide insight into the importance of miRNAs in the NED mechanism in PCa.<sup>60</sup> Analyses of NED by miRNA approaches would reveal novel mechanisms of NED and a therapeutic approach to neuroendocrine-differentiated PCa.

### MIRNAS ASSOCIATED WITH AR STATUS

PCa is initially AR-dependent, but it eventually acquires the AR-independent phenotype. Androgen signaling through AR is an important pathway for progression of PCa cells. During progression to CRPC, AR splice variant appears to increase in expression. AR-V7 is the most common AR splice variant without a ligand-binding domain, which is thought one of the mechanisms of progression of PCa to CRPC. It is known that PCa patients with AR-V7 in their circulating cancer cells do not respond to new hormonal agents (enzalutamide and abiraterone), because of androgen-independent proliferation of PCa cells with AR-V7.<sup>61</sup> Several miRNAs (*miR-21*, *-31*, *-34a*, *let-7c*, *-124*, *-205*, *-185*, *488\** and so on)<sup>9,23,62-66</sup> have been reported to regulate AR expression (both full length and AR-V7), and AR can regulate the expression of several miRs (*miR-21*, *-27a*, *-34*, *-125b*, *-221*, *-204*, *let-7*).<sup>9,60,67,68</sup>

Modulation of the AR transcriptional complex and AR co-repressor is an important mechanism in the response to androgen-deprivation therapy and eventual development of CRPC.<sup>69</sup> *miR-125b* is reported to directly target the AR co-repressor *NCOR2* and subsequently activate AR signaling.<sup>68</sup> AR inhibition has been shown to drive *miR-125b*, suggesting that androgen-deprivation therapy eventually results in the activation of AR by suppressing *NCOR* by *miR-125b*.<sup>68</sup>

A recent study has shown that *miR-21* and AR positively regulate each other and exert their oncogenic effects by inhibiting TGF $\beta$  receptor II (*TGFBR2*) expression.<sup>63</sup> They showed that *miR-21*/AR mediates its tumor-promoting function by attenuating TGF $\beta$ -mediated Smad2/3 activation, cell growth inhibition, cell migration and apoptosis in PCa.<sup>63</sup> *miR-124* is also reported to inhibit proliferation of PCa cells *in vitro* and sensitize them to inhibitors of AR signaling.<sup>61</sup>

*Let-7c* has been reported to suppress AR expression by degradation of *c-Myc*.<sup>62</sup> The *c-Myc* is an oncogenic transcription factor that is pathologically activated in many human malignancies including PCa, and *c-Myc* activity is known to induce androgen-independent PCa growth.<sup>70</sup> *Lin28* is a highly conserved RNA-binding protein known to be overexpressed in PCa. *Lin28* derepresses *c-Myc* by repressing *let-7c*, and *c-Myc* transcriptionally activates *Lin28*.<sup>71</sup> Thus, the *let-7c-Myc-Lin28* loop may have important roles in regulating AR expression and may help target-enhanced and hypersensitive ARs in advanced PCa.<sup>62</sup>

### MIRNAS AS PREDICTIVE BIOMARKERS IN PCa

In CRPC patients, PSA is not an appropriate marker to predict prognosis nor efficacy of treatment because poorly differentiated or neuroendocrine-differentiated PCa often show low levels of serum PSA. As a surrogate indicator, miRNAs can be attractive biomarkers because they are relatively stable in biological fluids, easy to measure and resistant to storage handling.<sup>9</sup> The aberrantly expressed miRNA levels in tumor tissue, serum or plasma, and urine can be a promising biomarker for PCa diagnosis, prognostic prediction or treatment efficacy.<sup>9</sup> In this review, we categorized miRNAs as predictive biomarkers by their origin, prostate tissue, serum or plasma, and urine.

The expression levels of miRNAs derived from prostate tissue have been analyzed and reported from multiple laboratories. Using radical prostatectomy specimens, *miR-21*, *-200a*, *-145*, *-30d*, *-301a*, *-449b* and *-182* have been mentioned as biomarkers to predict biochemical recurrence after prostatectomy.<sup>36,72-77</sup> Recently, combination of Gleason score and lymph node status with expression levels of *miR-4516* and *miR-601* has been reported to predict biochemical recurrence after post-prostatectomy salvage radiation therapy, supporting the use of miRNAs in clinically used predictive models.<sup>78</sup> Furthermore, Goto *et al.*<sup>10,53,79</sup> has reported that *miR-27b*, *miR-222* and *miR-452* could be potential biomarkers predicting progression time to CRPC.

A growing body of evidence indicates usefulness of circulating miRNAs in serum or plasma as biomarkers.<sup>80,81</sup> Circulating miRNAs can originate from tumor cells involved in tumor invasion or metastasis. Serum *miR-21*, *-375*, *-378\**, *141*, *-201*, *miR-200c*, *-423-3p* and *-210* have been reported as upregulated miRNAs in CRPC patients.<sup>82-88</sup> Furthermore, recent RNA sequencing of exosomal miRNAs in peripheral blood of CRPC patients indicates that higher expression of *miR-1290* and *miR-375* predicted poor overall survival.<sup>89</sup> Several reports describe the association of expression levels of circulating miRNAs and docetaxel chemotherapy for CRPC patients. Lin *et al.*<sup>90</sup> have revealed that non-responders to docetaxel therapy had high pre-docetaxel levels of serum *miR-200b*, and pre-docetaxel levels of serum *miR-200b* and post-docetaxel change in *miR-20a* levels were independent prognostic factors for overall survival. In other analysis, higher expression levels of serum *miR-21* was mentioned as a predictive factor for response to docetaxel.<sup>91</sup> Clinically monitoring these miRNAs would be useful to predict prognosis and sensitivity to therapeutic modality in CRPC patients.

Excretion of miRNAs in urine has been reported in several cancers including bladder cancer, renal cell carcinoma and PCa.<sup>92</sup> As for long non-coding RNA in urine, urinary prostate cancer antigen 3 (PCA3)

test is the most successfully developed marker in clinical use for PCa diagnosis.<sup>93</sup> Recently, it has been reported that miRNA levels in urine from PCa patients are significantly altered compared with that from benign prostate hypertrophy patients.<sup>94</sup> They showed that higher levels of urinary *miR-100* and *miR-200b* were effective parameters to detect the presence of PCa with PSA levels in the gray zone.<sup>94</sup> Other investigators identified *miR-205*, *-214*, *-1825* and *-484* as potential urinary biomarkers for PCa diagnosis.<sup>95,96</sup> At present, no reports describe association of urinary miRNA levels and risk of CRPC.

Combination of miRNAs with other conventional markers including Gleason score, clinical stage and PSA would be a suitable practical biomarker for patients to determine the most appropriate strategy to treat PCa and CRPC; however, studies with larger sample size are warranted to use miRNAs for biomarkers in the clinical settings.

### MIRNAS FOR TREATMENT OF PCA

Inhibition of oncogenic miRNAs or delivery of tumor-suppressive miRNAs could become a novel treatment strategy for PCa. As for tumor-suppressive miRNAs, adequate miRNA delivery system to the PCa tumors is required. However, the development of efficient *in vivo* miRNA delivery system has been challenging because of rapid degradation and excretion in serum condition and the lack of delivery system trapping miRNAs into the cancer cells.<sup>97</sup> A recent study has reported efficient miRNA delivery techniques using PCa-targeted nanoparticles (R11-SSPEI).<sup>98</sup> They showed R11-SSPEI/*miR-145* peptide inhibits intraperitoneal inoculated PC3 tumor growth *in vivo*. *miR-16*-conjugated atelocollagen has been shown to inhibit bone-metastatic human prostate xenograft growth in the mouse bone site *in vivo*.<sup>26</sup> Liposomal *miR-34* mimic (MRX34, Mirna Therapeutics Inc.) is under phase I clinical trial for liver cancer, lung cancer, malignant lymphoma, melanoma, multiple myeloma and renal cell carcinoma.<sup>99</sup> *miR-34* functions as tumor suppressor in PCa; thus, this *miR-34* delivery system will hopefully be useful for PCa in future. The optimization of the stability of miRNAs and improvement in delivery system of miRNAs are challenges for the future treatment of PCa.

### CONCLUSION

Accumulating evidence on the roles of miRNAs and the interactions between miRNAs and their target genes would promote a better understanding of PCa oncogenesis and castration resistance. Furthermore, determining roles of miRNAs that could be used as new diagnostic or predictive biomarkers would enable individualized therapeutic management for PCa patients. Elucidation of molecular mechanisms of PCa by miRNAs would help improve the therapeutic strategy of PCa.

### CONFLICT OF INTEREST

The authors declare no conflict of interest.

### ACKNOWLEDGEMENTS

This study was supported by the KAKENHI 26462430 (C).

- 4 Beer, T. M. & Tombal, B. Enzalutamide in metastatic prostate cancer before chemotherapy. *N. Engl. J. Med.* **371**, 1755–1756 (2014).
- 5 de Bono, J. S., Oudard, S., Ozguroglu, M., Hansen, S., Machiels, J. P., Kocak, I. et al. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. *Lancet* **376**, 1147–1154 (2010).
- 6 Shivelman, E., Beer, T. M. & Evans, C. P. Molecular pathways and targets in prostate cancer. *Oncotarget* **5**, 7217–7259 (2014).
- 7 Bartel, D. P. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* **116**, 281–297 (2004).
- 8 Coppola, V., De Maria, R. & Bonci, D. MicroRNAs and prostate cancer. *Endocr. Relat. Cancer*. **17**, F1–17 (2010).
- 9 Cannistraci, A., Di Pace, A. L., De Maria, R. & Bonci, D. MicroRNA as new tools for prostate cancer risk assessment and therapeutic intervention: results from clinical data set and patients' samples. *Biomed. Res. Int.* **2014**, 146170 (2014).
- 10 Goto, Y., Kojima, S., Nishikawa, R., Kurozumi, A., Kato, M., Enokida, H. et al. MicroRNA expression signature of castration-resistant prostate cancer: the microRNA-221/222 cluster functions as a tumour suppressor and disease progression marker. *Br. J. Cancer*. **113**, 1055–1065 (2015).
- 11 Volinia, S., Calin, G. A., Ambros, S., Cimmino, A., Petrocca, F. et al. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc. Natl Acad. Sci. USA* **103**, 2257–2261 (2006).
- 12 Schaefer, A., Jung, M., Mollenkopf, H. J., Wagner, I., Stephan, C., Jentzmik, F. et al. Diagnostic and prognostic implications of microRNA profiling in prostate carcinoma. *Int. J. Cancer* **126**, 1166–1176 (2010).
- 13 Fuse, M., Kojima, S., Enokida, H., Chiyomaru, T., Yoshino, H., Nohata, N. et al. Tumor suppressive microRNAs (miR-222 and miR-31) regulate molecular pathways based on microRNA expression signature in prostate cancer. *J. Hum. Genet.* **57**, 691–699 (2012).
- 14 Ottman, R., Nguyen, C., Lorch, R. & Chakrabarti, R. MicroRNA expressions associated with progression of prostate cancer cells to antiandrogen therapy resistance. *Mol. Cancer* **13**, 1 (2014).
- 15 Porkka, K. P., Pfeiffer, M. J., Waltering, K. K., Vessella, R. L., Tammela, T. L. & Visakorpi, T. MicroRNA expression profiling in prostate cancer. *Cancer Res.* **67**, 6130–6135 (2007).
- 16 Filella, X. & Foj, L. miRNAs as novel biomarkers in the management of prostate cancer. *Clin. Chem. Lab. Med.* (e-pub ahead of print 9 January 2016; doi:10.1515/cclm-2015-1073).
- 17 Ronnau, C. G., Verhaegh, G. W., Luna-Velez, M. V. & Schalken, J. A. Noncoding RNAs as novel biomarkers in prostate cancer. *Biomed. Res. Int.* **2014**, 591703 (2014).
- 18 Downing, S. R., Russell, P. J. & Jackson, P. Alterations of p53 are common in early stage prostate cancer. *Can. J. Urol.* **10**, 1924–1933 (2003).
- 19 Isaacs, W. B., Carter, B. S. & Ewing, C. M. Wild-type p53 suppresses growth of human prostate cancer cells containing mutant p53 alleles. *Cancer Res.* **51**, 4716–4720 (1991).
- 20 Navone, N. M., Troncoso, P., Pisters, L. L., Goodrow, T. L., Palmer, J. L., Nichols, W. W. et al. p53 protein accumulation and gene mutation in the progression of human prostate carcinoma. *J. Natl Cancer. Inst.* **85**, 1657–1669 (1993).
- 21 Shi, X. B., Xue, L., Ma, A. H., Tepper, C. G., Kung, H. J. & White, R. W. miR-125b promotes growth of prostate cancer xenograft tumor through targeting pro-apoptotic genes. *Prostate* **71**, 538–549 (2011).
- 22 Kojima, S., Inahara, M., Suzuki, H., Ichikawa, T. & Furuya, Y. Implications of insulin-like growth factor-I for prostate cancer therapies. *Int. J. Urol.* **16**, 161–167 (2009).
- 23 Sun, D., Layer, R., Mueller, A. C., Cichewicz, M. A., Negishi, M., Paschal, B. M. et al. Regulation of several androgen-induced genes through the repression of the miR-99a/let-7c/miR-125b-2 miRNA cluster in prostate cancer cells. *Oncogene* **33**, 1448–1457 (2014).
- 24 Melamed, J., Einhorn, J. M. & Ittmann, M. M. Allelic loss on chromosome 13q in human prostate carcinoma. *Clin. Cancer Res.* **3**, 1867–1872 (1997).
- 25 Bonci, D., Coppola, V., Musumeci, M., Addario, A., Giuffrida, R., Memeo, L. et al. The miR-15a-miR-16-1 cluster controls prostate cancer by targeting multiple oncogenic activities. *Nat. Med.* **14**, 1271–1277 (2008).
- 26 Takeshita, F., Patrawala, L., Osaki, M., Takahashi, R. U., Yamamoto, Y., Kosaka, N. et al. Systemic delivery of synthetic microRNA-16 inhibits the growth of metastatic prostate tumors via downregulation of multiple cell-cycle genes. *Mol. Ther.* **18**, 181–187 (2010).
- 27 Salmena, L., Carracedo, A. & Pandolfi, P. P. Tenets of PTEN tumor suppression. *Cell* **133**, 403–414 (2008).
- 28 Di Cristofano, A. & Pandolfi, P. P. The multiple roles of PTEN in tumor suppression. *Cell* **100**, 387–390 (2000).
- 29 Tian, L., Fang, Y. X., Xue, J. L. & Chen, J. Z. Four microRNAs promote prostate cell proliferation with regulation of PTEN and its downstream signals *in vitro*. *PLoS ONE* **8**, e75885 (2013).
- 30 Polisenio, L., Salmena, L., Riccardi, L., Fornari, A., Song, M. S., Hobbs, R. M. et al. Identification of the miR-106b~25 microRNA cluster as a proto-oncogenic PTEN-targeting intron that cooperates with its host gene MCM7 in transformation. *Sci. Signal.* **3**, ra29 (2010).
- 31 Si, M. L., Zhu, S., Wu, H., Lu, Z., Wu, F. & Mo, Y. Y. miR-21-mediated tumor growth. *Oncogene* **26**, 2799–2803 (2007).
- 32 Folin, M., Gandellini, P., Longoni, N., Profumo, V., Callari, M., Pennati, M. et al. miR-21: an oncomir on strike in prostate cancer. *Mol. Cancer*. **9**, 12 (2010).
- 33 Dong, B., Shi, Z., Wang, J., Wu, J., Yang, Z. & Fang, K. IL-6 inhibits the targeted modulation of PDCD4 by miR-21 in prostate cancer. *PLoS ONE* **10**, e0134366 (2015).

- 1 Siegel, R., Naishadham, D. & Jemal, A. Cancer statistics, 2013. *CA Cancer J. Clin.* **63**, 11–30 (2013).
- 2 James, N. D., Spears, M. R., Clarke, N. W., Dearnaley, D. P., De Bono, J. S., Gale, J. et al. Survival with newly diagnosed metastatic prostate cancer in the "docetaxel era": data from 917 patients in the control arm of the STAMPEDE Trial (MRC PR08, CRUK/06/019). *Eur. Urol.* **67**, 1028–1038 (2014).
- 3 Ryan, C. J., Smith, M. R., de Bono, J. S., Molina, A., Logothetis, C. J., de Souza, P. et al. Abiraterone in metastatic prostate cancer without previous chemotherapy. *N. Engl. J. Med.* **368**, 138–148 (2013).

- 34 Ribas, J. & Lupold, S. E. The transcriptional regulation of miR-21, its multiple transcripts, and their implication in prostate cancer. *Cell Cycle* **9**, 923–929 (2010).
- 35 Ribas, J., Ni, X., Haffner, M., Wentzel, E. A., Salmasi, A. H., Chowdhury, W. H. *et al*. miR-21: an androgen receptor-regulated microRNA that promotes hormone-dependent and hormone-independent prostate cancer growth. *Cancer Res.* **69**, 7165–7169 (2009).
- 36 Li, T., Li, R. S., Li, Y. H., Zhong, S., Chen, Y. Y., Zhang, C. M. *et al*. miR-21 as an independent biochemical recurrence predictor and potential therapeutic target for prostate cancer. *J. Urol.* **187**, 1466–1472 (2012).
- 37 Amankwah, E. K., Anegebe, E., Park, H., Pow-Sang, J., Hakam, A. & Park, J. Y. miR-21, miR-221 and miR-222 expression and prostate cancer recurrence among obese and non-obese cases. *Asian J. Androl.* **15**, 226–230 (2013).
- 38 Zheng, C., Yinghao, S. & Li, J. MiR-221 expression affects invasion potential of human prostate carcinoma cell lines by targeting DVL2. *Med. Oncol.* **29**, 815–822 (2012).
- 39 Galardi, S., Mercatelli, N., Giorda, E., Massalini, S., Frajese, G. V., Ciafre, S. A. *et al*. miR-221 and miR-222 expression affects the proliferation potential of human prostate carcinoma cell lines by targeting p27Kip1. *J. Biol. Chem.* **282**, 23716–23724 (2007).
- 40 Wang, L., Liu, C., Li, C., Xue, J., Zhao, S., Zhan, P. *et al*. Effects of microRNA-221-/222 on cell proliferation and apoptosis in prostate cancer cells. *Gene* **572**, 252–258 (2015).
- 41 Goto, Y., Kurozumi, A., Enokida, H., Ichikawa, T. & Seki, N. Functional significance of aberrantly expressed microRNAs in prostate cancer. *Int. J. Urol.* **22**, 242–252 (2015).
- 42 Wach, S., Nolte, E., Szczyrba, J., Stohr, R., Hartmann, A., Orntoft, T. *et al*. MicroRNA profiles of prostate carcinoma detected by multiplatform microRNA screening. *Int. J. Cancer.* **130**, 611–621 (2012).
- 43 Carlsson, J., Davidsson, S., Helenius, G., Karlsson, M., Lubovac, Z., Andren, O. *et al*. A miRNA expression signature that separates between normal and malignant prostate tissues. *Cancer Cell Int.* **11**, 14 (2011).
- 44 Ambs, S., Prueitt, R. L., Yi, M., Hudson, R. S., Howe, T. M., Petrocca, F. *et al*. Genomic profiling of microRNA and messenger RNA reveals deregulated microRNA expression in prostate cancer. *Cancer Res.* **68**, 6162–6170 (2008).
- 45 Szczyrba, J., Loprich, E., Wach, S., Jung, V., Unteregger, G., Barth, S. *et al*. The microRNA profile of prostate carcinoma obtained by deep sequencing. *Mol. Cancer Res.* **8**, 529–538 (2010).
- 46 Ozen, M., Creighton, C. J., Ozdemir, M. & Iltmann, M. Widespread deregulation of microRNA expression in human prostate cancer. *Oncogene* **27**, 1788–1793 (2008).
- 47 Ren, D., Wang, M., Guo, W., Zhao, X., Tu, X., Huang, S. *et al*. Wild-type p53 suppresses the epithelial-mesenchymal transition and stemness in PC-3 prostate cancer cells by modulating miR145. *Int. J. Oncol.* **42**, 1473–1481 (2013).
- 48 Fuse, M., Nohata, N., Kojima, S., Sakamoto, S., Chiyomaru, T., Kawakami, K. *et al*. Restoration of miR-145 expression suppresses cell proliferation, migration and invasion in prostate cancer by targeting FSCN1. *Int. J. Oncol.* **38**, 1093–1101 (2011).
- 49 Chiyomaru, T., Tatarano, S., Kawakami, K., Enokida, H., Yoshino, H., Nohata, N. *et al*. SWAP70, actin-binding protein, function as an oncogene targeting tumor-suppressive miR-145 in prostate cancer. *Prostate* **71**, 1559–1567 (2011).
- 50 Huang, S., Guo, W., Tang, Y., Ren, D., Zou, X. & Peng, X. miR-143 and miR-145 inhibit stem cell characteristics of PC-3 prostate cancer cells. *Oncol. Rep.* **28**, 1831–1837 (2012).
- 51 Ren, D., Wang, M., Guo, W., Huang, S., Wang, Z., Zhao, X. *et al*. Double-negative feedback loop between ZEB2 and miR-145 regulates epithelial-mesenchymal transition and stem cell properties in prostate cancer cells. *Cell Tissue Res.* **358**, 763–778 (2014).
- 52 Kojima, S., Enokida, H., Yoshino, H., Itesako, T., Chiyomaru, T., Kinoshita, T. *et al*. The tumor-suppressive microRNA-143/145 cluster inhibits cell migration and invasion by targeting GOLM1 in prostate cancer. *J. Hum. Genet.* **59**, 78–87 (2014).
- 53 Goto, Y., Kojima, S., Nishikawa, R., Enokida, H., Chiyomaru, T., Kinoshita, T. *et al*. The microRNA-23b/27b/24-1 cluster is a disease progression marker and tumor suppressor in prostate cancer. *Oncotarget* **5**, 7748–7759 (2014).
- 54 Visvader, J. E. & Lindeman, G. J. Cancer stem cells in solid tumours: accumulating evidence and unresolved questions. *Nat. Rev. Cancer* **8**, 755–768 (2008).
- 55 Patrawala, L., Calhoun, T., Schneider-Broussard, R., Li, H., Bhatia, B., Tang, S. *et al*. Highly purified CD44+ prostate cancer cells from xenograft human tumors are enriched in tumorigenic and metastatic progenitor cells. *Oncogene* **25**, 1696–1708 (2006).
- 56 DeSano, J. T. & Xu, L. MicroRNA regulation of cancer stem cells and therapeutic implications. *AAAPS J.* **11**, 682–692 (2009).
- 57 Li, J. & Lam, M. Reproducibility Project: Cancer, B. Registered report: the microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. *Elife* **4**, e06434 (2015).
- 58 Liu, C., Kelnar, K., Liu, B., Chen, X., Calhoun-Davis, T., Li, H. *et al*. The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. *Nat. Med.* **17**, 211–215 (2011).
- 59 Kong, D., Heath, E., Chen, W., Cher, M. L., Powell, I., Heilbrun, L. *et al*. Loss of let-7 up-regulates EZH2 in prostate cancer consistent with the acquisition of cancer stem cell signatures that are attenuated by BR-DIM. *PLoS ONE* **7**, e33729 (2012).
- 60 Ding, M., Lin, B., Li, T., Liu, Y., Li, Y., Zhou, X. *et al*. A dual yet opposite growth-regulating function of miR-204 and its target XRN1 in prostate adenocarcinoma cells and neuroendocrine-like prostate cancer cells. *Oncotarget* **6**, 7686–7700 (2015).
- 61 Shi, X. B., Ma, A. H., Xue, L., Li, M., Nguyen, H. G., Yang, J. C. *et al*. miR-124 and androgen receptor signaling inhibitors repress prostate cancer growth by downregulating androgen receptor splice variants, EZH2, and Src. *Cancer Res.* **75**, 5309–5317 (2015).
- 62 Nadiminty, N., Tummala, R., Lou, W., Zhu, Y., Zhang, J., Chen, X. *et al*. MicroRNA let-7c suppresses androgen receptor expression and activity via regulation of Myc expression in prostate cancer cells. *J. Biol. Chem.* **287**, 1527–1537 (2012).
- 63 Mishra, S., Deng, J. J., Gowda, P. S., Rao, M. K., Lin, C. L., Chen, C. L. *et al*. Androgen receptor and microRNA-21 axis downregulates transforming growth factor beta receptor II (TGFB2) expression in prostate cancer. *Oncogene* **33**, 4097–4106 (2014).
- 64 Lin, P. C., Chiu, Y. L., Banerjee, S., Park, K., Mosquera, J. M., Giannopoulou, E. *et al*. Epigenetic repression of miR-31 disrupts androgen receptor homeostasis and contributes to prostate cancer progression. *Cancer Res.* **73**, 1232–1244 (2013).
- 65 Qu, F., Cui, X., Hong, Y., Wang, J., Li, Y., Chen, L. *et al*. MicroRNA-185 suppresses proliferation, invasion, migration, and tumorigenicity of human prostate cancer cells through targeting androgen receptor. *Mol. Cell. Biochem.* **377**, 121–130 (2013).
- 66 Sikand, K., Slaibi, J. E., Singh, R., Slane, S. D. & Shukla, G. C. miR 488\* inhibits androgen receptor expression in prostate carcinoma cells. *Int. J. Cancer.* **129**, 810–819 (2011).
- 67 Ayub, S. G., Kaul, D. & Ayub, T. Microdissecting the role of microRNAs in the pathogenesis of prostate cancer. *Cancer Genet.* **208**, 289–302 (2015).
- 68 Yang, X., Bemis, L., Su, L. J., Gao, D. & Flaig, T. W. miR-125b regulation of androgen receptor signaling via modulation of the receptor complex co-repressor NCOR2. *Biores. Open Access* **1**, 55–62 (2012).
- 69 Hodgson, M. C., Astapova, I., Hollenberg, A. N. & Balk, S. P. Activity of androgen receptor antagonist bicalutamide in prostate cancer cells is independent of NCoR and SMRT corepressors. *Cancer Res.* **67**, 8388–8395 (2007).
- 70 Bernard, D., Pourtier-Manzanedo, A., Gil, J. & Beach, D. H. Myc confers androgen-independent prostate cancer cell growth. *J. Clin. Invest.* **112**, 1724–1731 (2003).
- 71 Chang, T. C., Zeitels, L. R., Hwang, H. W., Chivukula, R. R., Wentzel, E. A., Dews, M. *et al*. Lin-28B transactivation is necessary for Myc-mediated let-7 repression and proliferation. *Proc. Natl Acad. Sci. USA* **106**, 3384–3389 (2009).
- 72 Barron, N., Keenan, J., Gammell, P., Martinez, V. G., Freeman, A., Masters, J. R. *et al*. Biochemical relapse following radical prostatectomy and miR-200a levels in prostate cancer. *Prostate* **72**, 1193–1199 (2012).
- 73 Avgeris, M., Stravodimos, K., Fragoulis, E. G. & Scorilas, A. The loss of the tumour-suppressor miR-145 results in the shorter disease-free survival of prostate cancer patients. *Br. J. Cancer.* **108**, 2573–2581 (2013).
- 74 Kobayashi, N., Uemura, H., Nagahama, K., Okudela, K., Furuya, M., Ino, Y. *et al*. Identification of miR-30d as a novel prognostic maker of prostate cancer. *Oncotarget* **3**, 1455–1471 (2012).
- 75 Nam, R. K., Benatar, T., Wallis, C. J., Amemiya, Y., Yang, W., Garbens, A. *et al*. MiR-301a regulates E-cadherin expression and is predictive of prostate cancer recurrence. *Prostate* **76**, 869–884 (2016).
- 76 Mortensen, M. M., Hoyer, S., Orntoft, T. F., Sorensen, K. D., Dyrskjot, L. & Borre, M. High miR-449b expression in prostate cancer is associated with biochemical recurrence after radical prostatectomy. *BMC Cancer* **14**, 859 (2014).
- 77 Casanova-Salas, I., Rubio-Briones, J., Calatrava, A., Mancarella, C., Masia, E., Casanova, J. *et al*. Identification of miR-187 and miR-182 as biomarkers of early diagnosis and prognosis in patients with prostate cancer treated with radical prostatectomy. *J. Urol.* **192**, 252–259 (2014).
- 78 Bell, E. H., Kirste, S., Fleming, J. L., Stegmaier, P., Drendel, V., Mo, X. *et al*. A novel miRNA-based predictive model for biochemical failure following post-prostatectomy salvage radiation therapy. *PLoS ONE* **10**, e0118745 (2015).
- 79 Goto, Y., Kojima, S., Kurozumi, A., Kato, M., Okato, A., Matsushita, R. *et al*. Regulation of E3 ubiquitin ligase-1 (WWP1) by microRNA-452 inhibits cancer cell migration and invasion in prostate cancer. *Br. J. Cancer* **114**, 1135–1144 (2016).
- 80 Mitchell, P. S., Parkin, R. K., Kroh, E. M., Fritz, B. R., Wyman, S. K., Pogosova-Agdadjanyan, E. L. *et al*. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc. Natl Acad. Sci. USA* **105**, 10513–10518 (2008).
- 81 Chen, X., Ba, Y., Ma, L., Cai, X., Yin, Y., Wang, K. *et al*. Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res.* **18**, 997–1006 (2008).
- 82 Nguyen, H. C., Xie, W., Yang, M., Hsieh, C. L., Drouin, S., Lee, G. S. *et al*. Expression differences of circulating microRNAs in metastatic castration resistant prostate cancer and low-risk, localized prostate cancer. *Prostate* **73**, 346–354 (2013).
- 83 Zhang, H. L., Yang, L. F., Zhu, Y., Yao, X. D., Zhang, S. L., Dai, B. *et al*. Serum miRNA-21: elevated levels in patients with metastatic hormone-refractory prostate cancer and potential predictive factor for the efficacy of docetaxel-based chemotherapy. *Prostate* **71**, 326–331 (2011).
- 84 Cheng, H. H., Mitchell, P. S., Kroh, E. M., Dowell, A. E., Chery, L., Siddiqui, J. *et al*. Circulating microRNA profiling identifies a subset of metastatic prostate cancer patients with evidence of cancer-associated hypoxia. *PLoS ONE* **8**, e69239 (2013).
- 85 Brase, J. C., Johannes, M., Schlotmann, T., Falth, M., Haese, A., Steuber, T. *et al*. Circulating miRNAs are correlated with tumor progression in prostate cancer. *Int. J. Cancer* **128**, 608–616 (2011).
- 86 Shen, J., Hruby, G. W., McKiernan, J. M., Gurvich, I., Lipsky, M. J., Benson, M. C. *et al*. Dysregulation of circulating microRNAs and prediction of aggressive prostate cancer. *Prostate* **72**, 1469–1477 (2012).
- 87 Watahiki, A., Macfarlane, R. J., Gleave, M. E., Crea, F., Wang, Y., Helgason, C. D. *et al*. Plasma miRNAs as biomarkers to identify patients with castration-resistant metastatic prostate cancer. *Int. J. Mol. Sci.* **14**, 7757–7770 (2013).

- 88 Bryant, R. J., Pawlowski, T., Catto, J. W., Marsden, G., Vessella, R. L., Rhee, B. *et al.* Changes in circulating microRNA levels associated with prostate cancer. *Br. J. Cancer* **106**, 768–774 (2012).
- 89 Huang, X., Yuan, T., Liang, M., Du, M., Xia, S., Dittmar, R. *et al.* Exosomal miR-1290 and miR-375 as prognostic markers in castration-resistant prostate cancer. *Eur. Urol.* **67**, 33–41 (2015).
- 90 Lin, H. M., Castillo, L., Mahon, K. L., Chiam, K., Lee, B. Y., Nguyen, Q. *et al.* Circulating microRNAs are associated with docetaxel chemotherapy outcome in castration-resistant prostate cancer. *Br. J. Cancer* **110**, 2462–2471 (2014).
- 91 Zhang, G. M., Bao, C. Y., Wan, F. N., Cao, D. L., Qin, X. J., Zhang, H. L. *et al.* MicroRNA-302a suppresses tumor cell proliferation by inhibiting AKT in prostate cancer. *PLoS ONE* **10**, e0124410 (2015).
- 92 Huang, X., Liang, M., Dittmar, R. & Wang, L. Extracellular microRNAs in urologic malignancies: chances and challenges. *Int. J. Mol. Sci.* **14**, 14785–14799 (2013).
- 93 Groskopf, J., Aubin, S. M., Deras, I. L., Blase, A., Bodrug, S., Clark, C. *et al.* APTIMA PCA3 molecular urine test: development of a method to aid in the diagnosis of prostate cancer. *Clin. Chem.* **52**, 1089–1095 (2006).
- 94 Salido-Guadarrama, A. I., Morales-Montor, J. G., Rangel-Escareno, C., Langley, E., Peralta-Zaragoza, O. & Cruz Colin, J. L. *et al.* Urinary microRNA-based signature improves accuracy of detection of clinically relevant prostate cancer within the prostate-specific antigen grey zone. *Mol. Med. Rep.* **13**, 4549–4560 (2016).
- 95 Srivastava, A., Goldberger, H., Dimtchev, A., Ramalinga, M., Chijioke, J., Marian, C. *et al.* MicroRNA profiling in prostate cancer—the diagnostic potential of urinary miR-205 and miR-214. *PLoS ONE* **8**, e76994 (2013).
- 96 Haj-Ahmad, T. A., Abdalla, M. A. & Haj-Ahmad, Y. Potential urinary miRNA biomarker candidates for the accurate detection of prostate cancer among benign prostatic hyperplasia patients. *J. Cancer* **5**, 182–191 (2014).
- 97 Rothschild, S. I. microRNA therapies in cancer. *Mol. Cell. Ther.* **2**, 7 (2014).
- 98 Zhang, T., Xue, X., He, D. & Hsieh, J. T. A prostate cancer-targeted polyarginine-disulfide linked PEI nanocarrier for delivery of microRNA. *Cancer Lett.* **365**, 156–165 (2015).
- 99 Bader, A. G. miR-34 - a microRNA replacement therapy is headed to the clinic. *Front. Genet.* **3**, 120 (2012).
- 100 Dong, Q., Meng, P., Wang, T., Qin, W., Qin, W., Wang, F. *et al.* MicroRNA let-7a inhibits proliferation of human prostate cancer cells *in vitro* and *in vivo* by targeting E2F2 and CCND2. *PLoS ONE* **5**, e10147 (2010).
- 101 Tian, B., Huo, N., Li, M., Li, Y. & He, Z. let-7a and its target, insulin-like growth factor 1 receptor, are differentially expressed in recurrent prostate cancer. *Int. J. Mol. Med.* **36**, 1409–1416 (2015).
- 102 Shi, X. B., Xue, L., Yang, J., Ma, A. H., Zhao, J., Xu, M. *et al.* An androgen-regulated miRNA suppresses Bak1 expression and induces androgen-independent growth of prostate cancer cells. *Proc. Natl Acad. Sci. USA* **104**, 19983–19988 (2007).
- 103 Tao, J., Wu, D., Xu, B., Qian, W., Li, P., Lu, Q. *et al.* microRNA-133 inhibits cell proliferation, migration and invasion in prostate cancer cells by targeting the epidermal growth factor receptor. *Oncol. Rep.* **27**, 1967–1975 (2012).
- 104 Kojima, S., Chiyomaru, T., Kawakami, K., Yoshino, H., Enokida, H., Nohata, N. *et al.* Tumour suppressors miR-1 and miR-133a target the oncogenic function of purine nucleoside phosphorylase (PNP) in prostate cancer. *Br. J. Cancer* **106**, 405–413 (2012).
- 105 Xu, B., Niu, X., Zhang, X., Tao, J., Wu, D., Wang, Z. *et al.* miR-143 decreases prostate cancer cells proliferation and migration and enhances their sensitivity to docetaxel through suppression of KRAS. *Mol. Cell. Biochem.* **350**, 207–213 (2011).
- 106 Clape, C., Fritz, V., Henriquet, C., Apparailly, F., Fernandez, P. L., Iborra, F. *et al.* miR-143 interferes with ERK5 signaling, and abrogates prostate cancer progression in mice. *PLoS ONE* **4**, e7542 (2009).
- 107 Chu, H., Zhong, D., Tang, J., Li, J., Xue, Y., Tong, N. *et al.* A functional variant in miR-143 promoter contributes to prostate cancer risk. *Arch. Toxicol.* **90**, 403–414 (2016).
- 108 Ozen, M., Karatas, O. F., Gulluoglu, S., Bayrak, O. F., Sevil, S., Guzel, E. *et al.* Overexpression of miR-145-5p inhibits proliferation of prostate cancer cells and reduces SOX2 expression. *Cancer Invest.* **33**, 251–258 (2015).
- 109 Ngalame, N. N., Makia, N. L., Waalkes, M. P. & Tokar, E. J. Mitigation of arsenic-induced acquired cancer phenotype in prostate cancer stem cells by miR-143 restoration. *Toxicol Appl Pharmacol.* (e-pub ahead of print 22 December 2015; doi:10.1016/j.taap.2015.12.013).
- 110 Xie, S., Xie, Y., Zhang, Y. & Huang, Q. [Effects of miR-145 on the migration and invasion of prostate cancer PC3 cells by targeting DAB2]. *Yi Chuan.* **36**, 50–57 (2014).
- 111 Hart, M., Wach, S., Nolte, E., Szczyrba, J., Menon, R., Taubert, H. *et al.* The proto-oncogene ERG is a target of microRNA miR-145 in prostate cancer. *FEBS J.* **280**, 2105–2116 (2013).
- 112 Musumeci, M., Coppola, V., Addario, A., Patrizii, M., Maugeri-Sacca, M., Memeo, L. *et al.* Control of tumor and microenvironment cross-talk by miR-15a and miR-16 in prostate cancer. *Oncogene* **30**, 4231–4242 (2011).
- 113 Kong, D., Li, Y., Wang, Z., Banerjee, S., Ahmad, A., Kim, H. R. *et al.* miR-200 regulates PDGF-D-mediated epithelial-mesenchymal transition, adhesion, and invasion of prostate cancer cells. *Stem Cells* **27**, 1712–1721 (2009).
- 114 Banyard, J., Chung, I., Wilson, A. M., Vetter, G., Le Bechec, A., Bielenberg, D. R. *et al.* Regulation of epithelial plasticity by miR-424 and miR-200 in a new prostate cancer metastasis model. *Sci. Rep.* **3**, 3151 (2013).
- 115 Liu, Y. N., Yin, J. J., Abou-Kheir, W., Hynes, P. G., Casey, O. M., Fang, L. *et al.* MiR-1 and miR-200 inhibit EMT via Slug-dependent and tumorigenesis via Slug-independent mechanisms. *Oncogene* **32**, 296–306 (2013).
- 116 Wang, N., Li, Q., Feng, N. H., Cheng, G., Guan, Z. L., Wang, Y. *et al.* miR-205 is frequently downregulated in prostate cancer and acts as a tumor suppressor by inhibiting tumor growth. *Asian J. Androl.* **15**, 735–741 (2013).
- 117 Gandellini, P., Folini, M., Longoni, N., Pennati, M., Binda, M., Coecchia, M. *et al.* miR-205 Exerts tumor-suppressive functions in human prostate through down-regulation of protein kinase Cepsilon. *Cancer Res.* **69**, 2287–2295 (2009).
- 118 Bhatnagar, N., Li, X., Padi, S. K., Zhang, Q., Tang, M. S. & Guo, B. Downregulation of miR-205 and miR-31 confers resistance to chemotherapy-induced apoptosis in prostate cancer cells. *Cell Death Dis.* **1**, e105 (2010).
- 119 Hagman, Z., Hafidaddottir, B. S., Ceder, J. A., Larné, O., Bjartell, A., Lilja, H. *et al.* miR-205 negatively regulates the androgen receptor and is associated with adverse outcome of prostate cancer patients. *Br. J. Cancer* **108**, 1668–1676 (2013).
- 120 Wang, W., Liu, J. & Wu, Q. MiR-205 suppresses autophagy and enhances radio-sensitivity of prostate cancer cells by targeting TP53INP1. *Eur. Rev. Med. Pharmacol. Sci.* **20**, 92–100 (2016).
- 121 Reis, S. T., Pontes-Junior, J., Antunes, A. A., Dall'Oglio, M. F., Dip, N., Passerotti, C. *et al.* miR-21 may act as an oncomir by targeting RECK, a matrix metalloproteinase regulator, in prostate cancer. *BMC Urol.* **12**, 14 (2012).
- 122 Yang, X., Yang, Y., Gan, R., Zhao, L., Li, W., Zhou, H. *et al.* Down-regulation of miR-221 and miR-222 restrain prostate cancer cell proliferation and migration that is partly mediated by activation of SIRT1. *PLoS ONE* **9**, e98833 (2014).
- 123 Kneitz, B., Krebs, M., Kalogirou, C., Schubert, M., Joniau, S., van Poppel, H. *et al.* Survival in patients with high-risk prostate cancer is predicted by miR-221, which regulates proliferation, apoptosis, and invasion of prostate cancer cells by inhibiting IRF2 and SOCS3. *Cancer Res.* **74**, 2591–2603 (2014).
- 124 Xuan, H., Xue, W., Pan, J., Sha, J., Dong, B. & Huang, Y. Downregulation of miR-221, -30d, and -15a contributes to pathogenesis of prostate cancer by targeting Bmi-1. *Biochemistry (Mosc)* **80**, 276–283 (2015).
- 125 Majid, S., Dar, A. A., Saini, S., Arora, S., Shahryari, V., Zaman, M. S. *et al.* miR-23b represses proto-oncogene Src kinase and functions as methylation-silenced tumor suppressor with diagnostic and prognostic significance in prostate cancer. *Cancer Res.* **72**, 6435–6446 (2012).
- 126 Aghaee-Bakhtiari, S. H., Arefian, E., Naderi, M., Noorbakhsh, F., Nodouzi, V., Asgari, M. *et al.* MAPK and JAK/STAT pathways targeted by miR-23a and miR-23b in prostate cancer: computational and *in vitro* approaches. *Tumour Biol.* **36**, 4203–4212 (2015).
- 127 Ostling, P., Leivonen, S. K., Aakula, A., Kohonen, P., Makela, R., Hagman, Z. *et al.* Systematic analysis of microRNAs targeting the androgen receptor in prostate cancer cells. *Cancer Res.* **71**, 1956–1967 (2011).
- 128 Duan, K., Ge, Y. C., Zhang, X. P., Wu, S. Y., Feng, J. S., Chen, S. L. *et al.* miR-34a inhibits cell proliferation in prostate cancer by downregulation of SIRT1 expression. *Oncol. Lett.* **10**, 3223–3227 (2015).
- 129 Liang, H., Studach, L., Hullinger, R. L., Xie, J. & Andrisani, O. M. Down-regulation of RE-1 silencing transcription factor (REST) in advanced prostate cancer by hypoxia-induced miR-106b~25. *Exp. Cell Res.* **320**, 188–199 (2014).
- 130 Yamamura, S., Saini, S., Majid, S., Hirata, H., Ueno, K., Deng, G. *et al.* MicroRNA-34a modulates c-Myc transcriptional complexes to suppress malignancy in human prostate cancer cells. *PLoS ONE* **7**, e29722 (2012).
- 131 Kojima, K., Fujita, Y., Nozawa, Y., Deguchi, T. & Ito, M. MiR-34a attenuates paclitaxel-resistance of hormone-refractory prostate cancer PC3 cells through direct and indirect mechanisms. *Prostate* **70**, 1501–1512 (2010).