





















LETTER OPEN



ACUTE LYMPHOBLASTIC LEUKEMIA

Measurable residual disease quantification in adult patients with *KMT2A*-rearranged acute lymphoblastic leukemia

Thomas Burmeister^{1,13}, Aint-Steffen Ströh^{2,3,13}, Britta Kehden^{2,3}, Heiko Trautmann^{2,3}, Claus Meyer⁴, Rolf Marschalek⁴, Patrizia Larghero⁴, Stefan Schwartz⁵, Björn Steffen⁶, Bernd Spriewald⁷, Thomas Heinicke⁸, Nadja Jäkel⁹, Jörg Westermann¹, Kathrin Nachtkamp¹⁰, Andreas Viardot¹¹, Max S. Topp¹², Martin Neumann^{2,3}, Claudia D. Baldus^{2,3}, Nicola Gökbüget^{6,14} and Monika Brüggemann^{2,3,14}

© The Author(s) 2024

Leukemia; <https://doi.org/10.1038/s41375-024-02209-7>

TO THE EDITOR:

In adult ALL, 5–10% of patients show *KMT2A* translocations (*KMT2A* rearrangements) with only a few secondary alterations, implicating it as a leukemia-initiating factor [1, 2]. Approximately 95% of all fusions in adult ALL are *KMT2A::AFF1* or *KMT2A::MLLT1* [3]. *KMT2A*-rearranged adult ALL patients are generally considered high-risk and are treated with intensified therapy, including allogeneic hematopoietic stem cell transplantation (SCT) [4]. Current ALL treatment protocols are often guided by measurable residual disease (MRD)-based risk stratification [4–8], however, limited data are available regarding the prognostic value of MRD in adult ALL with *KMT2A* rearrangement. In infant *KMT2A*-rearranged ALL, more reliable MRD data were obtained using the individual *KMT2A* breakpoints as molecular MRD target as compared to *IG/TR* [6, 9–11], but no such comparisons have been made in adult ALL. We evaluated the impact of MRD on disease-free survival (DFS) and overall survival (OS) in a cohort of 156 *KMT2A*-rearranged adult patients and compared *IG/TR*- and *KMT2A*-based MRD levels in 46 patients.

In total, 769 bone marrow and/or peripheral blood samples from 193 adult ALL patients with *KMT2A* rearrangement (175 *KMT2A::AFF1*, 13 *KMT2A::MLLT1*, 1 *KMT2A::MLLT3*, 4 *KMT2A*+ unspecified) obtained between 2001 and 2021 were available for longitudinal MRD measurements. All patients were treated according to different protocols of the German Multicenter ALL (GMALL) study group and gave their informed consent to further scientific investigations on residual material. Patients with *KMT2A::AFF1* aged up to 55 years were assigned to the high-risk group and were candidates for SCT in first CR after consolidation I. Immunophenotyping and MRD measurement with real-time PCR based on *KMT2A* fusion genes

and clonal *IG/TR* gene rearrangements were performed in central laboratories as previously described [10, 11]. MRD measurements were interpreted according to EuroMRD guidelines [12]. MRD results were considered discordant if positivity/negativity discordance in the same sample was evidenced. For the evaluation of DFS and OS, MRD levels were compared at three different time points: end of induction I, after induction II/ pre-consolidation I, post-consolidation I/pre SCT (around week 16) (Fig. S2) [11]. MRD levels were classified as *molecular response* (MRD < 10⁻⁴ or negative), *molecular failure with low MRD* (≥10⁻⁴ and <10⁻²), and *high MRD* (≥10⁻²). Further statistical details are provided in the online supplement to this letter.

***KMT2A*-BASED VERSUS *IG/TR*-BASED MRD**

We analyzed 193 patients with *KMT2A*-rearranged ALL with median age at diagnosis of 42.5 years (18.0–76.8), and 63.0% being females. All 187 immunophenotypically characterized patients showed a CD10-negative B cell precursor ALL (146 *cyt*^{neg}, 41 *cyt*^{pos}). Parallel MRD data of both, *KMT2A* and *IG/TR*, were available for 46 patients, totaling 274 MRD data pairs from bone marrow and 99 from peripheral blood. Both methods show good agreement (Table S2; Fig. S1). 197/373 (52.8%) samples were MRD-negative with both methods, 84/373 (22.5%) were congruently positive within quantifiable range (QR), and 22/373 (5.9%) were positive below QR in both MRD targets (Fig. 1A). 18/373 (4.8%) were quantifiable MRD-positive only using *KMT2A*, whereas *IG/TR* MRD showed positivity below QR of the method, in 6/373 cases (1.6%) it was the other way around. The remaining 46/373 (13.0%) samples were classified as discordant, with 38/373 (10.2%) being *KMT2A*-rearranged and *IG/TR*^{neg}, with 24/46 samples showing quantifiable *KMT2A* MRD positivity. Only 8/373

¹Department of Hematology, Oncology and Tumor Immunology, CVK, Charité—Universitätsmedizin, corporate member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany. ²University Cancer Center Schleswig-Holstein (UCCSH), University Hospital Schleswig-Holstein, Kiel, Germany. ³Department of Medicine II, Hematology and Oncology, University Hospital Schleswig-Holstein, Kiel, Germany. ⁴Diagnostic Center of Acute Leukemia (DCAL), Institute of Pharmaceutical Biology, Goethe University, Frankfurt, Germany. ⁵Department of Hematology, Oncology and Tumor Immunology, CBF, Charité Universitätsmedizin, corporate member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany. ⁶Department of Medicine II, Goethe University, University Hospital, Frankfurt, Germany. ⁷University Hospital Erlangen, Department of Internal Medicine 5, Hematology and Oncology, Erlangen, Germany. ⁸Department of Hematology and Oncology, Otto-von-Guericke University Hospital, Magdeburg, Germany. ⁹Department of Hematology, Oncology, University Hospital, Halle/Saale, Germany. ¹⁰Department of Hematology, Oncology, University Hospital of Düsseldorf, Düsseldorf, Germany. ¹¹Department of Hematology, Oncology, University Hospital Ulm, Ulm, Germany. ¹²Department of Hematology, Oncology, University Hospital Würzburg, Würzburg, Germany. ¹³These authors contributed equally: Thomas Burmeister, Aint-Steffen Ströh. ¹⁴These authors jointly supervised this work: Nicola Gökbüget, Monika Brüggemann. ✉email: thomas.burmeister@charite.de

Received: 30 July 2023 Revised: 28 February 2024 Accepted: 28 February 2024

Published online: 22 March 2024

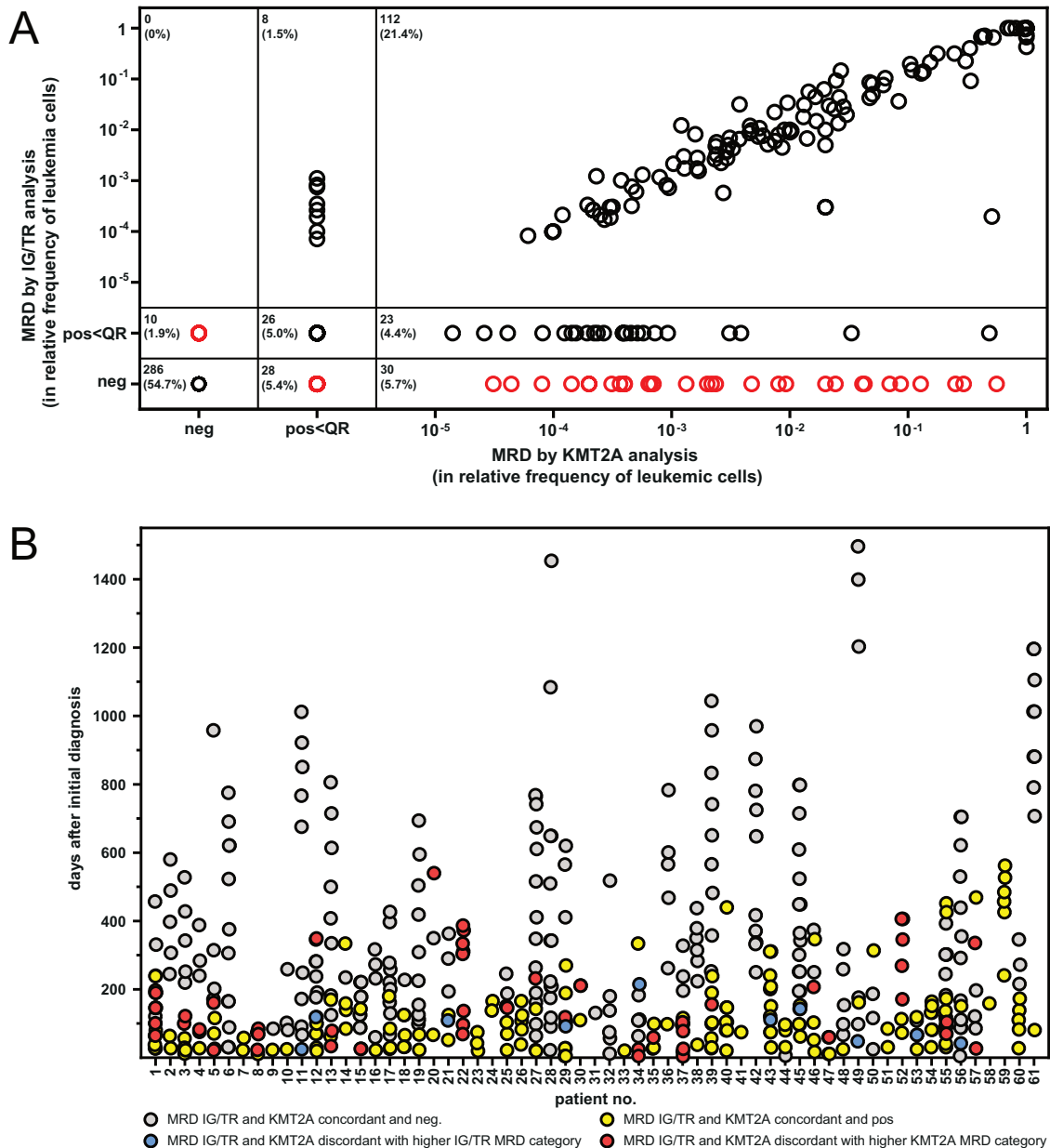


Fig. 1 Comparison of *KMT2A*-based and *IG/TR*-based MRD measurements. **A** Comparison of MRD levels with *KMT2A* and *IG/TR* targets in *KMT2A*-rearranged adult ALL patients. MRD measurements with data on both *KMT2A* and *IG/TR* were available from 46 patients totaling 373 sample pairs from peripheral blood or bone marrow aspirates. MRD levels were plotted against each other from negative (neg), positive (pos) < quantifiable range (QR), and quantifiable range in logarithmic format. Black circles represent MRD concordant samples and red circles discordant samples. **B** Comparison of *KMT2A* and *IG/TR* MRD levels over time. All MRD-levels ($n = 523$) with data on both *IG/TR* and *KMT2A* were sorted into four groups (gray color *IG/TR* and *KMT2A* MRD level concordant and negative (neg.), blue color *IG/TR* and *KMT2A* MRD level discordant with higher *IG/TR* MRD category, yellow color *IG/TR* and *KMT2A* MRD level concordant and positive, and red color *IG/TR* and *KMT2A* MRD level discordant with higher *KMT2A* MRD category) and plotted against days after initial diagnosis (ID).

(2.1%) were *KMT2A*^{neg} and *IG/TR*^{pos} ($P < 0.0001$), none of them showing quantifiable *IG/TR* MRD positivity (Table S1). Discordant samples with the higher *KMT2A* MRD were detected at least once in 15/46 (32.6%) patients during therapy and follow-up, whereas a higher *IG/TR* MRD was detected in 8/46 (17.4%) patients (Fig. 1B). These results are consistent with other studies on *KMT2A*-rearranged ALL, where *IG/TR* rearrangements at diagnosis were often oligo- or subclonal and underly clonal evolution [9]. Usage of subclonal *IG/TR* markers or a loss of the MRD marker due to RAG-mediated clonal evolution may lead to false negative results or underestimation of MRD values. In contrast, the *KMT2A* break cannot get lost because it is an early event and a leukemia-defining molecular hallmark.

PROGNOSTIC SIGNIFICANCE OF MRD

Data for DFS and OS in remission were available for 156/193 patients, with MRD being assessed using *IG/TR* and/or *KMT2A* at the end of induction I ($n = 140$), after induction II/ pre-consolidation I ($n = 149$) and after consolidation I ($n = 68$). After induction I, MRD levels did not predict outcome with 5-year DFS and OS ($P = 0.31$ and $P = 0.27$) (Fig. 2A+B). At pre-consolidation I, MRD levels did not predict outcome with 5-year DFS but patients with high MRD ($\geq 10^{-2}$) levels had a significantly poorer OS. 5-year OS was 62% (95% CI: 54 to 70), 59% (95% CI: 52 to 67), and 28% (95% CI: 11 to 45) for patients with molecular response, and molecular failure with low or high MRD ($P = 0.09$) (Fig. 2C+D).

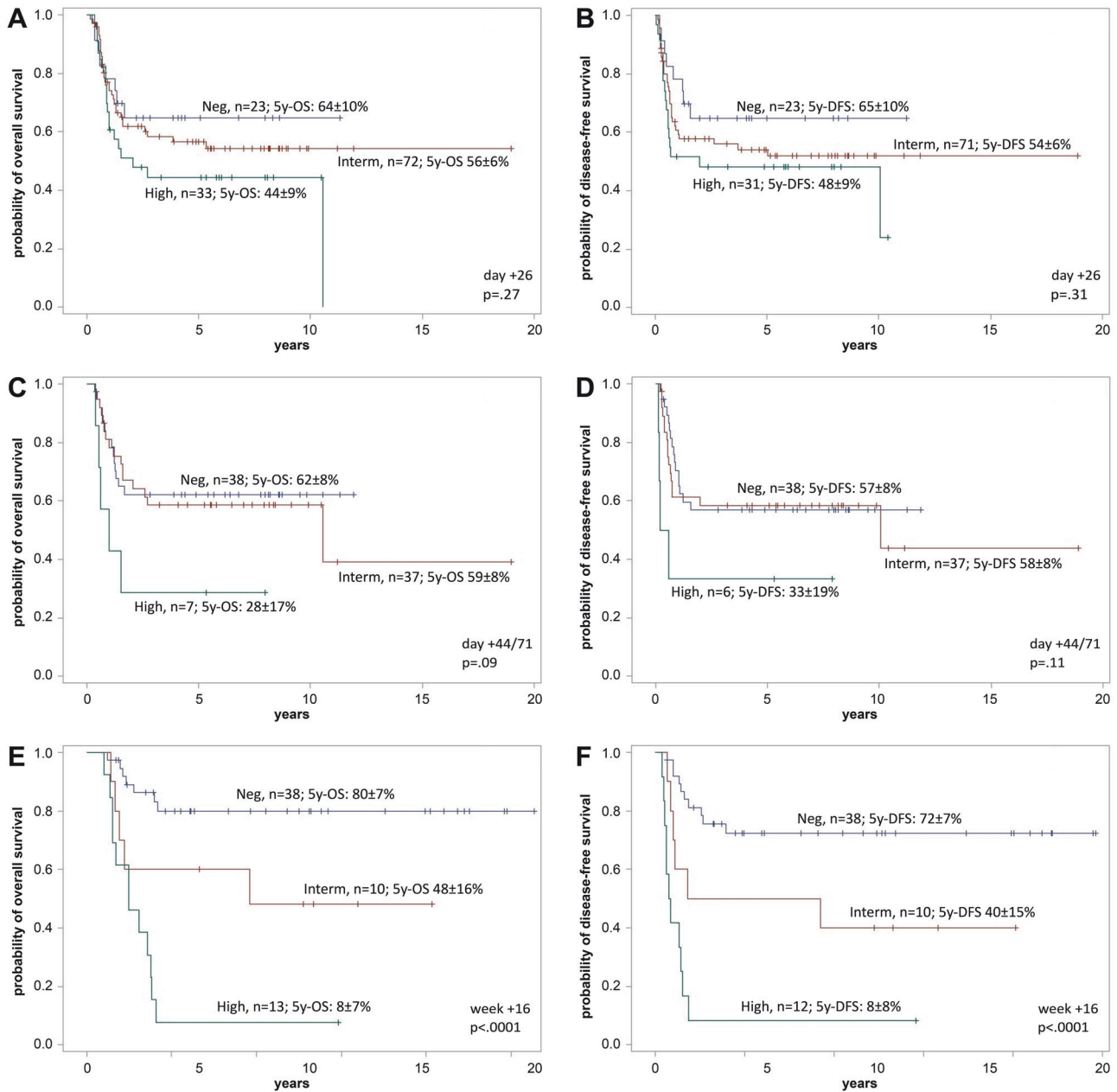


Fig. 2 Overall survival and disease-free survival of patients at different time points. Prognostic impact of measurable residual disease (MRD) levels at the end of induction I (A + B), pre-consolidation I (C + D), and post-consolidation I (E + F). Data shown by Kaplan–Meier estimates of overall survival (A, C, E) and disease-free survival (B, D, F). MRD results were classified as low (blue color; $<10^{-4}$), intermediate (red color; $<10^{-2}$), and high MRD levels (green color; $\geq 10^{-2}$). Information on DFS was not available for some patients (day+26: 3 pts., day+44/71: 1 pt., w +16: 1 pt.).

After consolidation I significant differences were found in both DFS and OS, and MRD levels predicted outcome with 5-year DFS of 72% (95% CI, 64 to 80), 40% (95% CI, 25 to 55), and 8% (95% CI, 0 to 16) and 5-year OS of 80% (95% CI, 73 to 88), 48% (95% CI, 32 to 64), and 8% (95% CI, 1 to 15) for patients with molecular response, and molecular failure with low ($\text{MRD} \geq 10^{-4}$ and $<10^{-2}$) and high ($\geq 10^{-2}$) MRD levels ($P < 0.0001$) (Fig. 2E+F). These findings demonstrate that high MRD levels at post-consolidation I in adult *KMT2A*-rearranged ALL are clearly unfavorable in terms of OS and DFS prior to SCT. Strikingly, early MRD after induction I was not predictive for treatment outcome, which contrasts with published data from other ALL molecular subgroups where early MRD has shown clear prognostic significance [5, 7, 11]. It is possible that this observation reflects the same mechanism that

has been described for infant *KMT2A*-rearranged ALL: In a study by Stutterheim et al. [13], MRD after induction was prognostically relevant only if followed by a lymphoid-style consolidation but not with a myeloid-style type consolidation. In our patient cohort allogeneic SCT was performed in the majority of patients (72%) which may abolish the prognostic effect of postinduction MRD response. However, patients with molecular failure prior to SCT still had poorer outcome. This supports the GMALL approach to offer a targeted therapy with blinatumomab to all patients with molecular failure after consolidation I to eradicate MRD before SCT [14]. However, patients with *KMT2A*-rearranged ALL occasionally show CD19 antigen loss after blinatumomab and blinatumomab may also be less effective than in non-*KMT2A*-rearranged ALL, due to lower CD19 expression.

MYELOID COEXPRESSION AND MRD RESPONSE

In the Interfant-06 study, patients with myeloid coexpression had significantly higher MRD levels at the end of induction and benefitted from subsequent myeloid-style consolidation [13]. In our cohort data were available in 96 patients for both, detailed immunophenotype and MRD. Expression of at least one myeloid marker (CD13, CD15, CD65, CD33) was detected in 77 (80.2%) patients. We observed no significant differences in MRD response at end of induction I, pre- or post-consolidation in patients with or without myeloid co-expression (Fig. S3; Table S3).

In conclusion, our study suggests that in adult *KMT2A*-rearranged ALL the *KMT2A* genomic fusion breakpoint has clear technical advantages as MRD target, as has also recently been reported by Kim et al. [15]. However, patient numbers in our study were too small to prove a clinical impact of MRD discordance between these two methods. The MRD status has a very strong prognostic value in DFS and OS post-consolidation I in a transplant-oriented regimen. The optimal therapy of patients with treatment failure or MRD persistence is under investigation. Particularly the term “myeloid-style therapy” needs to be defined more precisely, since most relevant compounds are also part of ALL therapy. More promise probably lies in the use of immunotherapies directed to lymphoid surface markers like CD19.

REFERENCES

- Andersson AK, Ma J, Wang J, Chen X, Gedman AL, Dang J, et al. The landscape of somatic mutations in infant MLL-rearranged acute lymphoblastic leukemias. *Nat Genet.* 2015;47:330–7.
- Aoki Y, Watanabe T, Saito Y, Kuroki Y, Hijikata A, Takagi M, et al. Identification of CD34+ and CD34- leukemia-initiating cells in MLL-rearranged human acute lymphoblastic leukemia. *Blood.* 2015;125:967–80.
- Burmeister T, Meyer C, Schwartz S, Hofmann J, Molkenin M, Kowarz E, et al. The MLL recombinome of adult CD10-negative B-cell precursor acute lymphoblastic leukemia: results from the GMALL study group. *Blood.* 2009;113:4011–5.
- Gökbuget N, Stelljes M, Viardot A, Nachtkamp K, Steffen B, Schneller F, et al. First results of the risk-adapted, MRD-stratified GMALL trial 08/2013 in 705 adults with newly diagnosed acute lymphoblastic leukemia/lymphoma (ALL/LBL). *Blood.* 2021;138:362.
- Beldjord K, Chevret S, Asnafi V, Huguet F, Boulland ML, Leguay T, et al. Onco-genetics and minimal residual disease are independent outcome predictors in adult patients with acute lymphoblastic leukemia. *Blood.* 2014;123:3739–49.
- van Dongen JJ, van der Velden VH, Brüggemann M, Orfao A. Minimal residual disease diagnostics in acute lymphoblastic leukemia: need for sensitive, fast, and standardized technologies. *Blood.* 2015;125:3996–4009.
- Gökbuget N, Kneba M, Raff T, Trautmann H, Bartram CR, Arnold R, et al. Adult patients with acute lymphoblastic leukemia and molecular failure display a poor prognosis and are candidates for stem cell transplantation and targeted therapies. *Blood.* 2012;120:1868–76.
- Bassan R, Spinelli O, Oldani E, Intermeoli T, Tosi M, Peruta B, et al. Improved risk classification for risk-specific therapy based on the molecular study of minimal residual disease (MRD) in adult acute lymphoblastic leukemia (ALL). *Blood.* 2009;113:4153–62.
- van der Velden VH, Corral L, Valsecchi MG, Jansen MW, De Lorenzo P, Cazzaniga G, et al. Prognostic significance of minimal residual disease in infants with acute lymphoblastic leukemia treated within the Interfant-99 protocol. *Leukemia.* 2009;23:1073–9.
- Burmeister T, Marschalek R, Schneider B, Meyer C, Gökbuget N, Schwartz S, et al. Monitoring minimal residual disease by quantification of genomic chromosomal breakpoint sequences in acute leukemias with MLL aberrations. *Leukemia.* 2006;20:451–7.
- Brüggemann M, Raff T, Flohr T, Gökbuget N, Nakao M, Droese J, et al. Clinical significance of minimal residual disease quantification in adult patients with standard-risk acute lymphoblastic leukemia. *Blood.* 2006;107:1116–23.
- van der Velden VH, Cazzaniga G, Schrauder A, Hancock J, Bader P, Panzer-Grümayer ER, et al. Analysis of minimal residual disease by Ig/TCR gene rearrangements: guidelines for interpretation of real-time quantitative PCR data. *Leukemia.* 2007;21:604–11.
- Stutterheim J, van der Sluis IM, de Lorenzo P, Alten J, Ancliffe P, Attarbaschi A, et al. Clinical implications of minimal residual disease detection in infants with *KMT2A*-rearranged acute lymphoblastic leukemia treated on the interfant-06 protocol. *J Clin Oncol.* 2021;39:652–62.
- Gökbuget N. MRD in adult Ph/BCR-ABL-negative ALL: how best to eradicate? *Hematology Am Soc Hematol Educ Program.* 2021;2021:718–25.
- Kim R, Bergugnat H, Pastoret C, Pasquier F, Raffoux E, Larcher L, et al. Genetic alterations and MRD refine risk assessment for *KMT2A*-rearranged B-cell precursor ALL in adults: a GRAALL study. *Blood.* 2023;142:1806–17.

ACKNOWLEDGEMENTS

We thank all participating clinics and patients of the GMALL study group. The technical assistance of Mara Molkenin, Daniela Gröger and Maïke Ipsen is highly appreciated. TB was supported by Deutsche José Carreras Leukämie-Stiftung grants 13 R/2016 and R10/37 f. This study was in part funded by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation)—“Clinician Scientist Program in Evolutionary Medicine” (GEPRIS project 4134905, to A-SS), and DFG project number 444949889 (KFO 5010/1 Clinical Research Unit ‘CATCH ALL’ to CB and MB).

AUTHOR CONTRIBUTIONS

TB, HT, CM, RM, SS, and MB performed experiments. A-SS, TB, BK, MN, NG, and MB analyzed results. NG, CM, RM, and SS provided relevant patient information for the study. TB, NG, and MB designed the research. A-SS drafted the first version of manuscript. All authors discussed the results and contributed to the final manuscript.

FUNDING

Open Access funding enabled and organized by Projekt DEAL.

COMPETING INTERESTS

MB is contracted to carry out research for Affimed, Amgen, Regeneron, the advisory board of Amgen, Incyte, Speaker bureau of Amgen, Janssen, Pfizer, Roche. SS is advisory board member of AMGEN and received honoraria as a speaker bureau member of AMGEN. CB is contracted to carry out research for Novartis, the advisory board of Amgen. TB received speakers’ honoraria from Novartis and Pfizer. The other authors declare that they have no conflict of interest.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41375-024-02209-7>.

Correspondence and requests for materials should be addressed to Thomas Burmeister.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2024